JC07 Rec'd PCT/PTO 07 DEC 2001

FORM PTO-1390 (REV. 9-2001) U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371

ATTORNEY'S DOCKET NUMBER: 2946-5181US

U.S. APPLICATION NO. (If known, see 37 CFR 1.5): 10/009109

INTERNATIONAL APPLICATION NO.	INTERNATIONAL FILING DATE	PRIORITY DATE CLAIMED
PCT/GB00/02100	9 June 2000 (09.06.00)	11 June 1999 (11.06.99)

TITLE OF INVENTION: ALLERGEN DETECTION

APPLICANT(S) FOR DO/EO/US: Ramin Pirzad

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (21) indicated below.
4. ☐ The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
   a. ☒ is attached hereto (required only if not communicated by the International Bureau).
   b. ☐ has been communicated by the International Bureau.
   c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
   a. ☐ is attached hereto.
   b. ☐ has been previously submitted under 35 U.S.C. 154(d)(4).
7. ☒ Amendments to the claims of the International Aplication under PCT Article 19 (35 U.S.C. 371(c)(3))
   a. ☐ are attached hereto (required only if not communicated by the International Bureau).
   b. ☐ have been communicated by the International Bureau.
   c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
   d. ☒ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371 (c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)) with Power of Attorney
10. ☐ An English lanugage translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11 to 20 below concern document(s) or information included:

11. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98 with Form PTO-1449 and citations
12. ☒ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A FIRST preliminary amendment.
14. ☐ A SECOND or SUBSEQUENT preliminary amendment.
15. ☐ A substitute specification.
16. ☐ A change of power of attorney and/or address letter.
17. ☐ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
18. ☐ A second copy of the published international application under 35 U.S.C. 154(d)(4).
19. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
20. ☒ Other items or information: International Preliminary Examination Report with Amended Sheets 16-20, claims 1-30
International Search Report
Notification of the Recording of a Change

NOTICE OF EXPRESS MAILING

Express Mail Label No.: EL 606649943US
Date of Deposit with USPS: December 7, 2001
Person making deposit: Blake Johnson

U.S. APPLICATION NO. (if known, see 37 CFR 1.5)	INTERNATIONAL APPLICATION NO	ATTORNEY'S DOCKET NUMBER
10/009109	PCT/GB00/02100	2946-5181US

21. ☒ The following fees are submitted:	CALCULATIONS PTO USE ONLY	
BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)):		
Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO **$1040.00**		
International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO **$890.00**		
International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO **$740.00**		
International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) **$710.00**	–	
International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) **$100.00**		
ENTER APPROPRIATE BASIC FEE AMOUNT =	$ 890.00	
Surcharge of **$130.00** for furnishing the oath or declaration later than ☐ 20 ☐ 30 months from the earliest claimed priority date (37 CFR 1.492(e)).	$	

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	
Total claims	30 - 20 =	10	x **$18.00**	$ 180.00
Independent claims	4 - 3 =	1	x **$84.00**	$ 84.00
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ **$280.00**	$
TOTAL OF ABOVE CALCULATIONS =				$ 1,154.00
☒ Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.			+	$ 577.00
SUBTOTAL =				$ 577.00
Processing fee of **$130.00** for furnishing the English translation later than ☐ 20 ☐ 30 months from the earliest claimed priority date (37 CFR 1.492(f)).				$
TOTAL NATIONAL FEE =				$ 577.00
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). **$40.00** per property +				$ 40.00
TOTAL FEES ENCLOSED =				$ 617.00
			Amount to be refunded:	$
			charged:	$

a. ☒ A check in the amount of $ 617.00 to cover the above fees is enclosed.

b. ☐ Please charge my Deposit Account No. _____ in the amount of $ _____ to cover the above fees. A duplicate copy of this sheet is enclosed.

c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 20-1469 . A duplicate copy of this sheet is enclosed.

d. ☐ Fees are to be charged to a credit card. **WARNING:** Information on this form may become public. **Credit card information should not be included on this form.** Provide credit card information and authorization on PTO-2038.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137 (a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

Joseph A. Walkowski
TRASKBRITT
230 South 500 East, Suite 300
P. O. Box 2550
Salt Lake City, Utah 84110

SIGNATURE

Joseph A. Walkowski
NAME

28,765
REGISTRATION NUMBER

Telephone: (801) 532-1922

PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:

Ramin Pirzad

Serial No.: Not Yet Assigned

Filed:

For: ALLERGEN DETECTION

Corresponding to: International Application No. PCT/GB00/02100

Examiner: Unknown

Group Art Unit: Unknown

Attorney Docket No.: 2946-5181US (P.6195)

NOTICE OF EXPRESS MAILING
Express Mail Mailing Label Number: EL 606649943US
Date of Deposit with USPS: December 7, 2001
Person making Deposit: Blake Johnson

PRELIMINARY AMENDMENT

Commissioner for Patents
Washington, D.C. 20231

Dear Sir:

Please preliminarily amend the above-identified application as follows, prior to calculating the filing fee and prior to examination on the merits. A marked-up version of the claims as amended herein is attached.

IN THE CLAIMS:

Please amend the claims as follows. A clean copy of all claims presently pending is set forth below. A marked-up version of each claim as amended herein is appended to this Amendment.

1. A method of determining allergen activity in dust, comprising:

providing a dust sample;

extracting from the dust sample at least one breakdown component of proteins or peptides;

reacting the extracted at least one breakdown component with a colorimetric amine detection reagent; and

quantitatively measuring the intensity of any resulting coloration, the allergen activity being proportional to the intensity of coloration.

2. A method according to claim 1, further comprising exposing the dust sample to a protease substrate, the protease substrate having immobilized thereon a protein or peptide on which protease in the dust sample may act.

3. A method according to claim 2, further comprising adding a protease inhibitor to the dust sample to suppress activity of a specific protease prior to exposure to the protease substrate.

4. A method according to claim 2, in which the protease substrate is protease specific, with only a specific protease being able to act on the protein or peptide immobilized on the substrate.

5. (Amended) A method according to claim 2, in which the protease substrate comprises a filter to facilitate extraction of mobile breakdown components of the protein or peptide immobilized on the protease substrate.

6. (Amended) A method according to claim 1, in which the breakdown components extracted from the dust sample include amines, amino acids or peptides present in the dust sample.

7. (Amended) A method according to claim 1, in which the colorimetric amine detection reagent is 2,4,6-trinitrobenzene sulphonic acid, (hereinafter referred to as TNBSA)

8. (Amended) A method according to claim 1, in which the at least one breakdown component is extracted by bringing the dust sample into contact with a surface active agent (surfactant).

9. A method according to claim 8, further comprising separating any dust sample solid residues from the surfactant prior to reacting with the colorimetric detection reagent.

10. (Amended) A method according to claim 8, in which the surfactant is an aqueous solution comprising sodium dodecyl sulphate.

11. A method according to claim 10, in which the aqueous solution is alkaline.

12. (Amended) A method according to claim 10, in which the aqueous solution further comprises sodium hydrogen carbonate.

13. (Amended) A method according to claim 1, in which the intensity of any resulting coloration is quantitatively measured by comparison with at least one reference color.

14. A method according to claim 13, in which different color references are selected to indicate at least three different kinds of allergen activity.

15. (Amended) A method according to claim 1, further comprising preserving the reaction mixture by using a stopping agent after a pre-selected incubation period.

16. A method of determining allergen activity in dust, comprising:

providing a dust sample;

providing a protease substrate, the protease substrate having immobilized thereon proteins or peptides labeled with a chromogenic substance;

exposing the protease substrate to the dust sample under conditions whereby a protease in the dust sample may act on the immobilized protein or peptide to produce mobile breakdown components labeled with the chromogenic substance;

and quantitatively measuring the intensity of any resulting coloration, the allergen activity being proportional to the intensity of the coloration.

17. A method according to claim 16, further comprising adding a protease inhibitor to the dust sample to suppress activity of a specific protease prior to exposure to the protease substrate.

18. A method according to claim 16, in which the protease substrate is protease specific, with only a specific protease being able to act on the proteins or peptides immobilized on the substrate.

19. (Amended) A method according to claim 16, in which the protease substrate comprises a filter to facilitate extraction of mobile breakdown components labeled with the chromogenic substance.

20. (Amended) A method according to claim 16, in which the intensity of any resulting coloration is quantitatively determined by comparison with at least one reference color.

21. Kit apparatus for use in a domestic environment for indicating allergen levels in dust, comprising a first chamber comprising a surfactant for extracting from a dust sample at least one breakdown component of proteins and peptides; a second chamber comprising a colorimetric amine detection reagent; means for quantitatively measuring the intensity of any coloration resulting from reacting the extract-containing surfactant and the colorimetric amine detection reagent; and means for indicating relative level of allergen activity in the dust sample based on the quantitative measurement.

22. Kit apparatus according to claim 21, further comprising a filter for filtering dust sample solid residues from the surfactant before reacting with the colorimetric amine detection reagent.

23. (Amended) Kit apparatus according to claim 21, in which one of the two chambers has the capacity to receive the contents of the other chamber.

24. Kit apparatus according to claim 23, in which the second chamber has the capacity to hold the colorimetric amine detection reagent and the surfactant.

25. (Amended) Kit apparatus according to claim 21, in which the quantitative measuring means comprises at least one color reference, against which the intensity of any coloration may be compared.

26. (Amended) Kit apparatus according to claim 21, in which the indicating means comprises a scale, which is linked to the intensity of any coloration measured.

27. (Amended) Kit apparatus according to claim 21, further comprising a third chamber comprising a stopping reagent to limit the reaction between the extract-containing surfactant and the colorimetric amine detection reagent.

28. (Amended) Kit apparatus according to claim 21, in which the colorimetric amine detection reagent is 2,4,6-trinitrobenzene sulphonic acid.

29. Apparatus for use in determining allergen levels in a dust sample, comprising a protease substrate having immobilized thereon proteins or peptides labeled with a chromogenic substance, whereby any protease in the dust sample may act on the immobilized proteins or peptides to produce mobile breakdown components labeled with the chromogenic substance.

30. (Amended) Apparatus according to claim 29, in which proteins labeled with the chromogenic substance comprise azo-albumin.

REMARKS

No new matter has been added. The Applicant requests entry of the foregoing Amendment prior to calculation of the filing fee and examination of the application on the merits. An early Office Action on the merits is respectfully solicited.

Correspondence of Claims to Claims in the PCT Application

For the convenience of the Examiner, Applicant herein notes that the claims as amended herein correspond in substance to those as amended during Chapter II proceedings under the PCT, and as appended to the International Preliminary Examination Report (IPER) as Amended Sheets 16 through 20 including thereon claims 1 through 30 (copy of IPER enclosed with Amended Sheets 16 through 20). This Preliminary Amendment removes multiple dependencies from a number of the claims and, in the case of claim 30, corrects a minor dependency error and a single grammatical error. Otherwise, the claims are identical to claims 1 through 30 as amended during Chapter II proceedings under the PCT. The amendments made herein do not narrow the scope of the claims, nor were they introduced to avoid any prior art known to Applicant.

Respectfully submitted,

Joseph A. Walkowski
Registration No. 28,765
Attorney for Applicant
TRASKBRITT, PC
P. O. Box 2550
Salt Lake City, Utah 84110-2550
Telephone: (801) 532-1922

JAW\csk
Date: December 7, 2001
N:\2946\5181\Preliminary Amendment.wpd

MARKED-UP VERSION SHOWING CHANGES MADE

5. (Amended) A method according to claim 2, [3 or 4,] in which the protease substrate comprises a filter to facilitate extraction of mobile breakdown components of the protein or peptide immobilized on the protease substrate.

6. (Amended) A method according to [any one of claims 1 to 5] claim 1, in which the breakdown components extracted from the dust sample include amines, amino acids or peptides present in the dust sample.

7. (Amended) A method according to [any one of claims 1 to 6] claim 1, in which the colorimetric amine detection reagent is 2,4,6-trinitrobenzene sulphonic acid, (hereinafter referred to as TNBSA)

8. (Amended) A method according to [any one of claims 1 to 7] claim 1, in which the at least one breakdown component is extracted by bringing the dust sample into contact with a surface active agent (surfactant).

10. (Amended) A method according to claim 8 [or 9], in which the surfactant is an aqueous solution comprising sodium dodecyl sulphate.

12. (Amended) A method according to claim 10 [or 11], in which the aqueous solution further comprises sodium hydrogen carbonate.

13. (Amended) A method according to [any one of claims 1 to 12] claim 1, in which the intensity of any resulting coloration is quantitatively measured by comparison with at least one reference color.

15. (Amended) A method according to [any one of claims 1 to 14] claim 1, further comprising preserving the reaction mixture by using a stopping agent after a pre-selected incubation period.

19. (Amended) A method according to claim 16, [17 or 18,] in which the protease substrate comprises a filter to facilitate extraction of mobile breakdown components labeled with the chromogenic substance.

20. (Amended) A method according to [any one of claims 16 to 19] claim 16, in which the intensity of any resulting coloration is quantitatively determined by comparison with at least one reference color.

23. (Amended) Kit apparatus according to claim 21 [or 22], in which one of the two chambers has the capacity to receive the contents of the other chamber.

25. (Amended) Kit apparatus according to [any one of claims 21 to 24] claim 21, in which the quantitative measuring means comprises at least one color reference, against which the intensity of any coloration may be compared.

26. (Amended) Kit apparatus according to [any one of claims 21 to 24] claim 21, in which the indicating means comprises a scale, which is linked to the intensity of any coloration measured.

27. (Amended) Kit apparatus according to [any one of claims 21 to 24] claim 21, further comprising a third chamber comprising a stopping reagent to limit the reaction between the extract-containing surfactant and the colorimetric amine detection reagent.

28. (Amended) Kit apparatus according to [any one of claims 21 to 27] claim 21, in which the colorimetric amine detection reagent is 2,4,6-trinitrobenzene sulphonic acid.

30. (Amended) Apparatus according to claim [20] 29, in which proteins labeled with the chromogenic [the] substance comprise azo-albumin.

TITLE: ALLERGEN DETECTION

TECHNICAL FIELD

The present invention relates to allergen detection, and more particularly to a method and apparatus for indicating allergen levels in dust samples.

BACKGROUND ART

It is estimated that up to 80% of the dust particles illuminated by incident sunlight and made visible to the naked eye in a domestic environment are derived from skin. In a warm environment, dust mites feed on skin-derived dust particles, breaking it down by using proteases in their digestive system. Such proteases are found in not insignificant levels in dust mite faeces, and it is now established that it is excreted proteases which act as allergens to individuals who are liable to have an allergic response to house dust. Concentrations of excreted protease are found in relatively high levels in carpets, bedding, pillows and mattresses, all of which provide a suitable environment for dust mites to thrive.

Dust mites are not the only source of proteases

found in house dust. For example, proteases from cockroaches are also a source of allergens. Furthermore it is possible that proteases from cat saliva become airborne as the saliva dries, for example, on the cat's fur. It is likely that such proteases also act as allergens to individuals who are allergic to house dust.

It is known to test house dust in order to determine quantitatively levels of the house dust mite allergen. According to one patent, US 4,806,490, a dust sample is suspended in an aqueous-alcoholic alkali metal hydroxide solution to dissolve or leach out aromatic compounds such as guanine excreted by dust mites, and the resulting solution is mixed with an aromatic diazo compound. A reaction between the aromatic diazo compound and certain excreted aromatic compounds in the solution produces a colour change, with the intensity of the new colour being indicative of the level of excreted proteases in the house dust.

DISCLOSURE OF THE INVENTION

According to a first aspect of the present invention, there is provided a method of determining allergen activity in dust, comprising: providing a dust sample; extracting from the dust sample at least one breakdown component of proteins or peptides; reacting the extracted at least one breakdown component with a colorimetric amine detection reagent; and determining or quantitatively measuring the intensity of any resulting coloration, the allergen activity being proportioned to

the intensity of coloration.

The present applicant has appreciated that in addition to proteases, dust mites excrete the by-products of skin breakdown, including amine compounds, amino acids and relatively small chain peptides, e.g., glycylglycine. In part, the present invention is directed to detecting some of the more abundant, and in some cases chemically less complex, by-products to give an indication of the allergen concentration, rather than targeting one specific compound (e.g., guanine) or type of compounds (e.g., aromatic compounds). This will enable individuals to test particular environments, e.g., individual rooms in a domestic situation to establish that environment's propensity for inducing an allergic response.

The method may further comprise exposing the dust sample to a protease substrate, the protease substrate having immobilised thereon proteins or peptides on which protease in the dust sample may act. The protease substrate may comprise a physical support, such as a matrix or membrane. Thus, in this way, the breakdown components of proteins or peptides will at least in part be generated *in situ*. This may be useful for increasing the concentration of such components, and hence improving subsequent quantitative coloration intensity measurements. If this technique is employed, the exposure time of the dust sample to the protease substrate may need to be controlled (e.g. set at 15 minutes). It is to be noted that in such a process the allergen is

effectively being measured directly.

The method may further comprise adding a protease inhibitor to the dust sample to suppress activity of a specific protease prior to exposure to the protease substrate. In certain circumstances, it may be necessary to distinguish between dust mite protease and another protease (e.g. from cockroaches), since an individual may be more allergic to one than the other. Differentiation between the types of proteases present in the dust sample can be achieved by differential inhibition of certain specific proteases which may be present. For example, serine protease inhibitors may be used to inhibit specifically serine proteases. The serine protease inhibitors may be selected from the group consisting of organophosphates (e.g. diisopropylphosphofluoridate), sulphonyl fluorides (e.g. phenylmethylsulphonyl fluoride), coumarins (e.g. 3,4-dichloroisocoumarin) and peptide/protein inhibitors (e.g. peptide boronic acids and aprotinin, respectively). The use of serine protease inhibitors would allow dust mite allergens (e.g. cysteine proteases) to be detected more readily. On the other hand cysteine protease inhibitors may be used if dust mite allergens were to be excluded from the test. The cysteine protease inhibitors may be selected from the group consisting of peptide diazomethanes (e.g. z-Phe-Ala-CHN2), and peptide epoxides (e.g. E-64 and its derivatives), cystatins.

In one embodiment of the method, the protease

substrate is protease specific, with only a specific protease being able to act on the protein or peptide immobilised on the substrate. In this way, the protease substrate may be chosen to target a specific protease which may be present in the dust sample. If the protease is present in the sample, the specific proteins or peptides immobilised on the substrate will be broken down for subsequent detection. On the other hand, if the specific protease is absent, the proteins or peptides will remain intact and immobilised on the substrate.

The protease substrate may comprise a filter to facilitate extraction of mobile breakdown components of the proteins or peptides immobilised on the protease substrate. The filter may even act as a barrier to the passage of proteases therethrough. The breakdown components extracted from the dust sample may include amines, amino acids or peptides either from the dust sample or from the protease substrate.

The colorimetric amine detection reagent may be 2,4,6-trinitrobenzene sulphonic acid (hereinafter TNBSA).

The at least one breakdown component may be extracted by bringing the dust sample into contact with a surface active agent (surfactant). Any dust sample solid residues may be separated from the surfactant prior to reacting with the colorimetric amine detection reagent. The surfactant may be an aqueous solution comprising sodium dodecyl sulphate, possibly present in an amount of about 5 wt%. The aqueous solution may be alkaline and

may also comprise sodium hydrogen carbonate. The dust sample solid residues may be separated by filtration. Removing the solid residues facilitates accurate determination of the intensity of any coloration by reducing the amount of opaque material in the solution.

The intensity of any resulting coloration may be quantitatively determined by comparison with at least one reference colour. The comparison may be with a plurality of different colour references, each selected from the spectrum of colours or range of colour hues attainable. The different colour references may be selected to indicate at least three different kinds of allergen activity, perhaps corresponding to a macroscopic gradation such as low, medium and high activity.

The reaction mixture may be preserved by using a stopping agent, e.g., hydrochloric acid, after a pre-selected incubation or dwell time, e.g., about 2 minutes.

In order to give reproducible results, the dust sample may be of a predetermined size, e.g., by weight or by volume. The dust sample may be collected by a suction device, perhaps over a predetermined area or time. Variations in the dust sample size may be tolerated since the method represents a gross contamination test, so exact measurements of the dust samples are not necessarily essential.

In accordance with a second aspect of the invention, there is provided a method of determining allergen activity in dust, comprising: providing a dust sample;

providing a protease substrate, the protease substrate having immobilised thereon proteins or peptides labelled with a chromogenic substance; exposing the protease substrate to the dust sample under conditions whereby any protease in the dust sample may act on the immobilised protein or peptide to produce mobile breakdown components labelled with the chromogenic substance; and quantitatively measuring the intensity of any resulting coloration, the allergen activity being proportional to the intensity of the coloration.

The method may further comprise adding a protease inhibitor to the dust sample to suppress activity of a specific protease prior to exposure to the protease substrate. As before, this will enable the specific protease to be excluded from becoming actively involved in the test, allowing other protease - perhaps present in lower concentrations - to be evaluated. For example, the inhibitor may be a cysteine protease inhibitor if protease allergens other than those from dust mites are to be evaluated.

In another embodiment of the invention, the protease substrate may be protease specific, with only a specific protease being able to act on the proteins or peptides immobilised on the substrate. In this way, the test may be tailored to evaluate a specific protease, regardless of whether different kinds of protease are present in the dust sample. For example, synthetic substrates with 4-nitroaniline and 2-naphthylamine (chromophores) can be

used to distinguish between metaloproteases and aspartic proteases on the one hand (e.g. from cockroaches) and serine and cysteine proteases on the other hand (e.g. from dust mites).

The protease substrate may comprise a filter to facilitate extraction of mobile breakdown components labelled with the chromogenic substance. The filter may act as a barrier to all molecules which are larger than mobile breakdown components labelled with the chromogenic substance.

An example of a protein labelled with a chromogenic substance is azo-albumin. When reacted with a suitable protease, an azo-dye is released.

In accordance with a third aspect of the present invention, there is provided a method of determining allergen activity in dust, comprising: providing a dust sample; extracting from the dust sample at least one component selected from the group consisting of aliphatic amines and aliphatic amino acids; determining the relative concentration of the extracted at least one component; and providing an indication of allergen activity in dependence upon the relative concentration determined.

The relative concentration may be determined by employing a colour indicator sensitive to aliphatic amines and amino acids. The colour indicator may comprise TNBSA.

Any by-products of skin breakdown, particularly

aliphatic amines and aliphatic amino acids, present in the dust sample may be linked to dust mite activity. The higher the levels of the by-products in the dust sample, the higher the dust mite activity may be assumed to be. High levels of dust mite activity will produce a correspondingly high amount of protease - the allergens which are largely responsible for providing the allergic reaction to house dust in certain individuals.

In accordance with a fourth aspect of the invention, there is provided a method of determining allergen-generation propensity in dust, comprising: providing a dust sample; exposing the dust sample to a protease able to break down proteins or peptides in human skin cells; reacting the exposed dust sample with a colorimetric amine detection reagent; and quantitatively measuring the intensity of any resulting coloration, the allergen-generation propensity being proportioned to the intensity of the coloration.

An individual may want to evaluate a dust sample to see whether it might support a high level of dust mite activity, even before the allergen levels have built up to significant, detectable levels. If the dust sample contains relatively high levels of human skin cells, the protease supplied will produce breakdown components which will react with the reagent and thereby be detected by colour evaluation. By containing relatively high levels of human skin cells, the dust sampled could in theory support high concentrations of dust mite. Such

information may be a useful warning to those individuals who are allergic to dust mite protease.

The colorimetric amine detection reagent may be 2,4,6-trinitrobenzene sulphonic acid. The intensity of any resulting coloration may be quantitatively measured by comparison with at least one reference colour.

In accordance with another aspect of the present invention, there is provided apparatus for use in a domestic environment for determining indicating allergen levels. The apparatus may comprise a kit comprising a first chamber comprising a surfactant for extracting from a dust sample at least one breakdown component and of proteins and peptides; a second chamber comprising a colorimetric amine detection reagent; means for quantitatively measuring the intensity of any coloration resulting from reacting the extract-containing surfactant and the colorimetric amine detection reagent; and means for indicating relative level of allergen activity in the dust sample based on the quantitative measurement.

The apparatus may further comprise a filter for filtering dust sample solid residues from the surfactant before reacting with the colorimetric amine detection reagent, which may be TNBSA. One of the two chambers may have the capacity to receive the contents of the other chamber. Preferably, the second chamber has the capacity to hold the colorimetric amine detection apparatus and the surfactant.

The quantitative measuring means may comprise at

least one colour reference, against which the colour of the solution may be compared. The indicating means may comprise a scale, e.g., low, medium and high activity, which is linked to the intensity of any coloration measured. For example, if the colour of the solution is determined by eye as being about the same as the colour reference, this could correspond to medium allergen activity. Divergence either side of the colour reference would then correspond to low or high activity as appropriate.

The apparatus may further comprise a third chamber comprising a stopping reagent to limit the reaction between the extract-containing surfactant and colorimetric amine detection reagent, e.g. TNBSA.

BRIEF DESCRIPTION OF THE DRAWINGS

An embodiment of the invention will now be described with reference to the accompanying drawings, in which:

Figure 1 shows schematically apparatus for determining dust mite activity in accordance with the present invention;

Figure 2 shows schematically the use of apparatus shown in Figure 1;

Figure 3 is a flow chart illustrating one method of determining allergen levels according to the invention; and

Figure 4 is a flow chart illustrating another method embodying the invention.

MODES OF CARRYING OUT THE INVENTION

The apparatus 10 of figure 1 comprises three parts: an upper part 12 which contains in a first chamber 14 0.10 litres of a 0.1M solution of sodium hydrogen carbonate containing 5 wt% of sodium dodecyl sulphate; a middle part 16 which is a snug but sliding fit in both the upper part 12 and the remaining part; and a lower part 18 which contains a tablet of TNBSA and a stopping reagent of 1.0M hydrochloric acid. The solution in the first chamber 14 is sealed in the upper part 12 by a frangible seal 20. The middle part 16 comprises a filter 22 above which is provided a cup 24 for receiving a dust sample. The middle part 16 has a leading profile 26 which is pointed to facilitate breaking the frangible seal 20. A second chamber 27 is formed by the middle and lower parts. The lower part 18 includes a frangible seal 28 disposed between the tablet of TNBSA and the stopping reagent which is sealed in a third chamber 29.

The use of the apparatus 10 is now described in stages with reference to figure 2:

Stage 1 A sample of dust of predetermined size is placed in cup 24.

Stage 2 The middle part 16 is inserted into the upper part 12, such that the profile 26 ruptures the seal 20.

Stage 3 The solution in the first chamber comes into contact with the dust sample. Any chemicals including amines, amino acids and peptides present in the

dust sample are extracted and pass through filter 22 and into the second chamber where they come into contact with the tablet of TNBSA.

Stage 4 After about 2 minutes, the middle part 16 is pushed far enough into the lower part 18 to rupture seal 28, enabling the stopping reagent in the third chamber 29 to prevent further reaction. The colour of the resulting solution is compared with a colour key which is calibrated to give an indication of the level (e.g., low, medium or high) of dust mite activity in the dust sample.

Example

A dust sample was collected from an old mattress (where dust mite activity may be expected to be high), and a blank sample and test samples of GlycylGlycine in varying concentrations (20-200 micro-grams) were used as controls. The dust, blank and test samples were washed with 0.1M $NaHCO_3$ 0.5M NaCl (pH 8.3)and then tested with TNBSA of various concentrations e.g. diluted to 1 part in 10, 1 part in 50 and 1 part in 100. It was found that a dilution of 1 part in 50 was the optimum dilution for sensitivity and blank colour. Using such a dilution, the experiment yielded visual results for both the dust and all test samples, but not the blank sample. The visual results could then be assessed and compared to give an indication of dust mite activity in the old mattress.

The method used in the example may be summarised and developed with reference to Figure 3. A dust sample is

provided at step 50, possibly by using a suction device to collect dust from furniture or carpets. A protease inhibitor (e.g. serine protease inhibitor) is added at step 52) to enable a particular protease (e.g. cysteine protease) to be targeted. Next, at step 54, the dust sample is exposed to a protease substrate which is exposed to a protease substrate which is susceptible to the proteases present. Protein or peptide breakdown components from the dust sample or protease substrate are then extracted at 56 and are reacted at 58 with the colorimetric amine detection reagent (TNBSA). The presence of free amino groups causes an orange-coloured product, the intensity of which is measured at 60 to give an indication of allergen levels.

Instead of using a protease inhibitor (step 52), the protease substrate may be selected to be protease specific. In other words, the protease substrate may contain proteins or peptides which require the presence of specific proteases under evaluation before yielding detectable breakdown components.

An alternative method is illustrated in Figure 4, and again starts with the provision of a dust sample (again step 50). A specific protease substrate is provided at 70; the substrate having immobilised thereon proteins or peptides which require specific proteases before yielding breakdown components. The immobilised proteins or peptides are also labelled with a chromogenic substance. At step 72, the substrate is exposed to the

dust sample. The presence of the specific proteases in the sample will break down the immobilised proteins or peptides, releasing the chromogenic substance, causing coloration of the solution. The intensity of the coloration is measured at 74 to give an indication of allergen levels.

Instead of using a protease-specific substrate (step 70), the protease substrate may be non-specific, but still labelled with the chromogenic substance. If a specific protease is still to be targeted, this may be accomplished using protease inhibitors (step 52 of Figure 3).

CLAIMS

1. A method of determining allergen activity in dust, comprising:

providing a dust sample;

extracting from the dust sample at least one breakdown component of proteins or peptides;

reacting the extracted at least one breakdown component with a colorimetric amine detection reagent; and

quantitatively measuring the intensity of any resulting coloration, the allergen activity being proportional to the intensity of coloration.

2. A method according to claim 1, further comprising exposing the dust sample to a protease substrate, the protease substrate having immobilised thereon a protein or peptide on which protease in the dust sample may act.

3. A method according to claim 2, further comprising adding a protease inhibitor to the dust sample to suppress activity of a specific protease prior to exposure to the protease substrate.

4. A method according to claim 2, in which the protease substrate is protease specific, with only a specific protease being able to act on the protein or peptide immobilised on the substrate.

5. A method according to claim 2,3 or 4, in which the protease substrate comprises a filter to facilitate extraction of mobile breakdown components of the protein or peptide immobilised on the protease substrate.

6. A method according to any one of claims 1 to 5, in

which the breakdown components extracted from the dust sample include amines, amino acids or peptides present in the dust sample.

7. A method according to any one of claims 1 to 6, in which the colorimetric amine detection reagent is 2,4,6-trinitrobenzene sulphonic acid, (hereinafter referred to as TNBSA)

8. A method according to any one of claims 1 to 7, in which the at least one breakdown component is extracted by bringing the dust sample into contact with a surface active agent (surfactant).

9. A method according to claim 8, further comprising separating any dust sample solid residues from the surfactant prior to reacting with the colorimetric detection reagent.

10. A method according to claim 8 or 9, in which the surfactant is an aqueous solution comprising sodium dodecyl sulphate.

11. A method according to claim 10, in which the aqueous solution is alkaline.

12. A method according to claim 10 or 11, in which the aqueous solution further comprises sodium hydrogen carbonate.

13. A method according to any one of claims 1 to 12, in which the intensity of any resulting coloration is quantitatively measured by comparison with at least one reference colour.

14. A method according to claim 13, in which different

colour references are selected to indicate at least three different kinds of allergen activity.

15. A method according to any one of claims 1 to 14, further comprising preserving the reaction mixture by using a stopping agent after a pre-selected incubation period.

16. A method of determining allergen activity in dust, comprising:

providing a dust sample;

providing a protease substrate, the protease substrate having immobilised thereon proteins or peptides labelled with a chromogenic substance;

exposing the protease substrate to the dust sample under conditions whereby a protease in the dust sample may act on the immobilised protein or peptide to produce mobile breakdown components labelled with the chromogenic substance;

and quantitatively measuring the intensity of any resulting coloration, the allergen activity being proportional to the intensity of the coloration.

17. A method according to claim 16, further comprising adding a protease inhibitor to the dust sample to suppress activity of a specific protease prior to exposure to the protease substrate.

18. A method according to claim 16, in which the protease substrate is protease specific, with only a specific protease being able to act on the proteins or peptides immobilised on the substrate.

19. A method according to claim 16,17 or 18, in which the protease substrate comprises a filter to facilitate extraction of mobile breakdown components labelled with the chromogenic substance.

20. A method according to any one of claims 16 to 19, in which the intensity of any resulting coloration is quantitatively determined by comparison with at least one reference colour.

21. Kit apparatus for use in a domestic environment for indicating allergen levels in dust, comprising a first chamber comprising a surfactant for extracting from a dust sample at least one breakdown component of proteins and peptides; a second chamber comprising a colorimetric amine detection reagent; means for quantitatively measuring the intensity of any coloration resulting from reacting the extract-containing surfactant and the colorimetric amine detection reagent; and means for indicating relative level of allergen activity in the dust sample based on the quantitative measurement.

22. Kit apparatus according to claim 21, further comprising a filter for filtering dust sample solid residues from the surfactant before reacting with the colorimetric amine detection reagent.

23. Kit apparatus according to claim 21 or 22, in which one of the two chambers has the capacity to receive the contents of the other chamber.

24. Kit apparatus according to claim 23, in which the second chamber has the capacity to hold the colorimetric

amine detection reagent and the surfactant.

25. Kit apparatus according to any one of claims 21 to 24, in which the quantitative measuring means comprises at least one colour reference, against which the intensity of any coloration may be compared.

26. Kit apparatus according to any one of claims 21 to 24, in which the indicating means comprises a scale, which is linked to the intensity of any coloration measured.

27. Kit apparatus according to any one of claims 21 to 24, further comprising a third chamber comprising a stopping reagent to limit the reaction between the extract-containing surfactant and the colorimetric amine detection reagent.

28. Kit apparatus according to any one of claims 21 to 27, in which the colorimetric amine detection reagent is 2,4,6-trinitrobenzene sulphonic acid.

29. Apparatus for use in determining allergen levels in a dust sample, comprising a protease substrate having immobilised thereon proteins or peptides labelled with a chromogenic substance, whereby any protease in the dust sample may act on the immobilised proteins or peptides to produce mobile breakdown components labelled with the chromogenic substance.

30. Apparatus according to claim 20, in which proteins labelled with chromogenic the substance comprise azo-albumin.

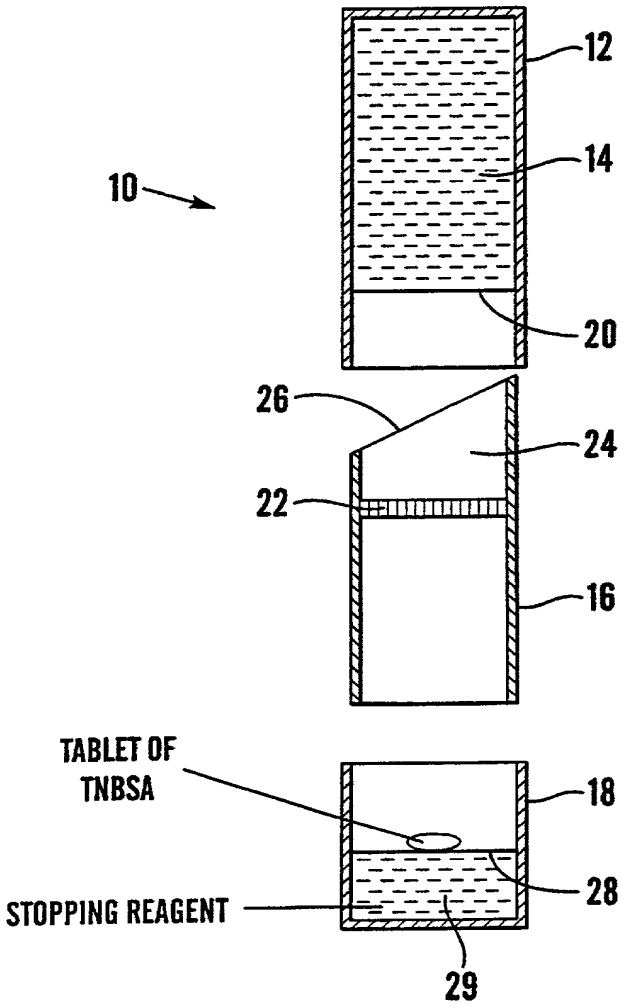
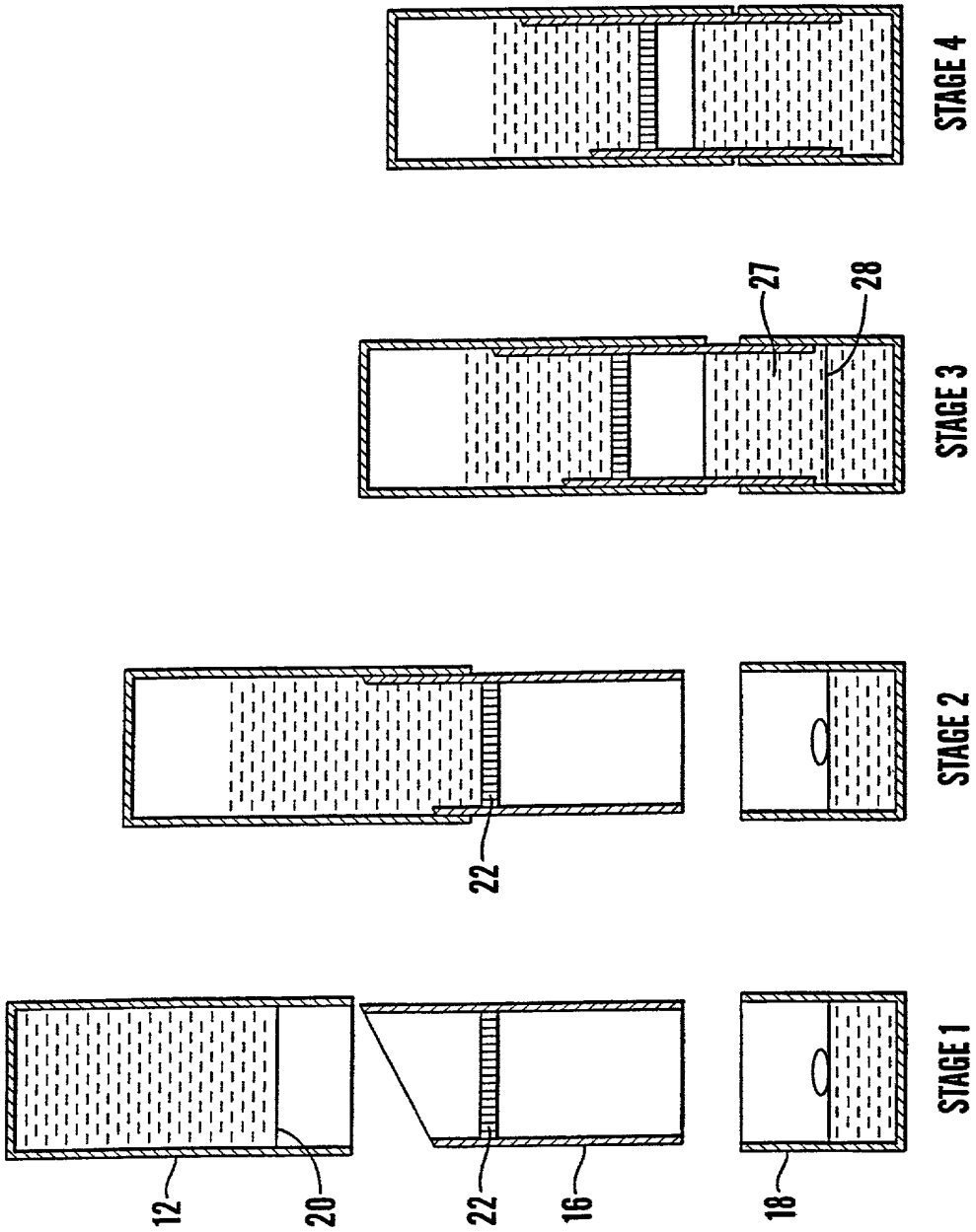

10
12
14
20
26
24
22
16
TABLET OF TNBSA
18
28
STOPPING REAGENT
29
SAMPLE CUP HOLDING APPROXIMATELY 1g OF DUST

Fig. 1

Fig.2



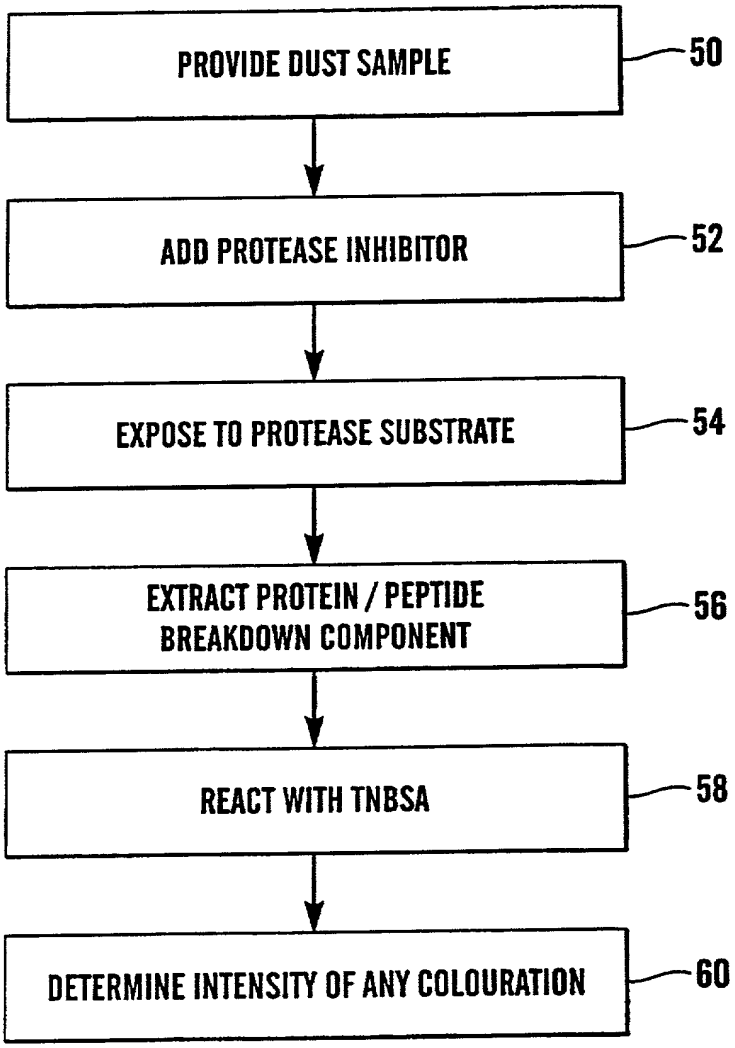


Fig.3

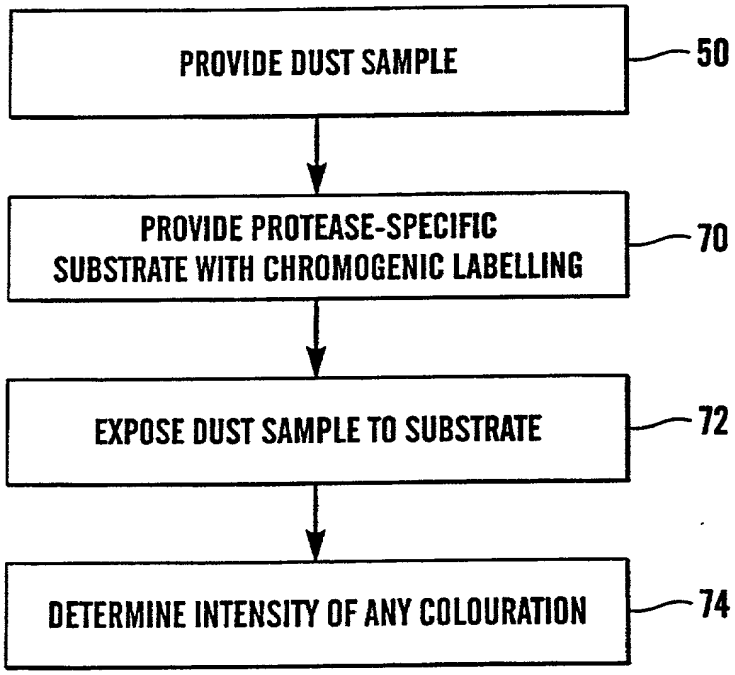

PROVIDE DUST SAMPLE
50
PROVIDE PROTEASE-SPECIFIC SUBSTRATE WITH CHROMOGENIC LABELLING
70
EXPOSE DUST SAMPLE TO SUBSTRATE
72
DETERMINE INTENSITY OF ANY COLOURATION
74

Fig.4

Attorney Docket No. 5181US

DECLARATION FOR PATENT APPLICATION (WITH POWER OF ATTORNEY)

As an inventor named below or on any attached continuation page, I hereby declare that:

My residence, post office address and citizenship are as stated next to my name.

I believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled "ALLERGEN DETECTION," the specification of which (check one):

☐ is attached hereto
☒ was filed on _____ as United States application serial no. _____ and was amended on _____.
☒ was filed on June 9, 2000, as PCT international application no. PCT/GB00/02100 and was amended under PCT Article 19 on _____.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the U.S. Patent and Trademark Office all information known to me to be material to the patentability of the subject matter claimed in this application, as "materiality" is defined in Title 37, Code of Federal Regulations § 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate or § 365(a) of any PCT international application(s) designating at least one country other than the United States of America listed below and on any attached continuation page and have also identified below and on any attached continuation page any foreign application for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America having a filing date before that of the application(s) on which priority is claimed.

Prior foreign/PCT application(s):

(number)	(country)	(day/month/year filed)	Priority Claimed Yes	No
9913487.6	GB	11 June 1999	X	

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) or § 365(c) of PCT international application(s) designating the United States of America listed below and on any attached continuation page and, insofar as the subject matter of each of the claims of this application is not disclosed in any such prior application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose to the U.S. Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations § 1.56 which became available between the filing date of such prior application and the national or PCT international filing date of this application:

(application serial no.)	(filing date)	(status - pending, patented or abandoned)
PCT/GB00/02100	9 June 2000	Pending

I hereby claim the benefit under Title 35, United States Code, § 119(e) of any United States provisional application(s) listed below:

(provisional application no.)	(filing date)

I hereby appoint the following Registered Practitioners to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith:

David V. Trask, Reg. No. 22,012
Joseph A. Walkowski, Reg. No. 28,765
Allen C. Turner, Reg. No. 33,041
Bruce E. Hayden, Reg. No. 35,539
Devin R. Jensen, Reg. No. 44,805
Shawn G. Hansen, Reg. No. 42,627
Andrew F. Nilles, Reg. No. 47,825

William S. Britt, Reg. No. 20,969
James R. Duzan, Reg. No. 28,393
Edgar R. Cataxinos, Reg. No. 39,931
Brick G. Power, Reg No. 38,581
Krista Weber Powell, Reg. No. 47,867
Bretton L. Crockett, Reg. No. 44,632
Katherine A. Hamer, Reg. No. 47,628

Laurence B. Bond, Reg. No. 30,549
H. Dickson Burton, Reg. No. 48,396
Kent S. Burningham, Reg. No. 30,453
Paul C. Oestreich, Reg. No. 44,983
David L. Stott, Reg. No. 43,937
Bradley B. Jensen, Reg. No. 46,801

Address all correspondence to:

Joseph A. Walkowski
TRASKBRITT
P.O. Box 2550
Salt Lake City, Utah 84110
Telephone No. (801) 532-1922

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of sole inventor: RAMIN PIRZAD

Inventor's signature R. Pirzad Date 23/11/01

Residence: 40 Nursery Gardens, St. Ives, Cambridgeshire PE21 3NL, GREAT BRITAIN GBN
Citizenship: British
Post Office Address: 40 Nursery Gardens, St. Ives, Cambridgeshire PE21 3NL, GREAT BRITAIN